Dylan Ross Weaver

Current Lab: https://weaverlab.owlstown.net/

Lab: Phillips Science 209

Doctor of Philosophy - Physics

EDUCATION

University of Missouri

Columbia, MO, United States

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August 2019 - May 2024

Office: Phillips Science 239

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Dissertation Title: Probing Membrane Protein Dynamics and Interactions with Lipids at the Single Molecule Level

Advisor: Dr. Gavin M. King

University of Missouri

Master of Science - Physics; GPA: 3.8

Columbia, MO, United States

August 2019 - July 2022

Graduate Minor: College Teaching

 $\textbf{\textit{Courses:}}\ \ \textit{Nonlinear Dynamics, Structural Biology and Biophysics for Life Sciences, Computational Biological Physics, Single Molecule}$ 

Biophysics, Techniques of Teaching College Physics, College Science Teaching

Murray State University

Bachelor of Science, Magna Cum Laude - Physics, Mathematics; GPA: 3.65

Murray, KY, United States August 2015 - May 2019

TEACHING APPOINTMENTS

University of Wisconsin Eau Claire

On Campus

Assistant Professor

Aug 2024 - Present

• Lecture and Laboratory Instructor: Implemented active learning strategies in courses for both majors and non-majors.

University of Missouri

On Campus and Online

Graduate Teaching Assistant: In Person and Online

Aug 2019 - May 2022

- $\circ \ \textbf{Instructor} : \ \textbf{Fulfilled the Minor in College Teaching program as an instructor for PHYS 1200 Everyday Wonders}$
- o **Head Teaching Assistant**: Primary contact for all TA's serving in Calculus Based Physics II (PHYS 2760)
- Lab/Discussion Teaching Assistant: Laboratory TA for Calculus Based Physics II, as well as discussion sections for Calc. and Alg. based Physics I/II. This includes regular office hours and grading exams and lab reports.

**Murray State University** 

On Campus

Undergraduate Teaching Assistant

Aug 2015 - May 2019

- Online Grading Coordinator: Implemented online software for TA's to streamline grading tasks each week. This includes developing rubrics for lab reports in all introductory Physics courses.
- Laboratory Teaching Assistant: Primary and/or secondary TA for Calc. and Alg. based Physics I/II courses, as well as Introductory Astronomy.

# TEACHING EXPERIENCE BY COURSE TITLE

## University of Wisconsin Eau Claire

Assistant Professor

- PHYS 100 Introduction to Physical Science: A conceptual study of physics using ideas of force and energy to understand motion, properties of matter, sound, light, and electricity and magnetism. Enrollment: 40-60 students
- PHYS 232 University Physics II: A continuation of Physics 231 (University Physics I), including the study of electricity, magnetism, and optics. Enrollment: 40-50 students
- PHYS 365 Theoretical Mechanics: Newton's laws, accelerated frames, central-force orbits, angular momentum of systems, coupled oscillations, generalized coordinates, and Lagrange's equations. Enrollment: 10-20 students

## University of Missouri

Instructor of Record

• PHYS 1200 - Everyday Wonders: How Ordinary Things Work: A conceptual course that focuses on how one can use the principles of Physics to describe processes in everyday life. Students completed online assignments along with weekly lab meetings. Average Enrollment: 20-30 students

### Teaching Awards

• Harry B. Hammond Award for Excellence in Undergraduate Teaching - May 2022

## CURRENT RESEARCH: SINGLE MOLECULE BIOLOGICAL PHYSICS

- Biological Atomic Force Microscopy (AFM): Membrane proteins make up 30% per of proteins in cells, performing various critical functions. Many techniques provide relevant biochemical information, but most only provide static conformations. AFM can be a complementary tool in studying the dynamic structure of these proteins in physiologically relevant conditions and real time. The method can also be used to study how certain peptides disrupt membrane morphology. Along with molecular-scale (10 Å) lateral resolution and ~1 Å vertical resolution, trace/retrace kymographs can be used to achieve sub-100 ms temporal resolution.
- Single Molecule Force Spectroscopy: Studying the interactions between certain peptides and lipid membranes is essential for probing various cellular processes and mechanisms. The challenge in studying these interactions resides in not only the complexity of the system, but the length and time scales that the interactions take place in. Through AFM-based force spectroscopy, we can probe the dissociation forces and free energy profiles of various peptides as they negotiate a membrane. Applying SMFS to peptide-lipid interaction can help to quantify amino-acid contributions with single residue precision, as well as effects due to the hydrophobicity of certain peptides.

## DETAILED RESEARCH EXPERIENCE

## University of Wisconsin Eau Claire

Assistant Professor

Aug 2024 - Present

- Imaging Lipid Membranes in Presence of Maximin 3: Maximin peptides are secreted by the Chinese red-belly toad *Bombina maxima*, and possess antimicrobial activity. Through imaging and spectroscopy, one can shed light on the mechanism of action for membrane disruption. One can also determine various mechanical properties of the membrane film, such as the line tension, in the presence and absence of maximins.
- The Role of Hydrophobicity on Peptide-Lipid Interactions: Numerous hydrophobicity scales have been utilized to describe which amino acids readily partition into a membrane or prefer to be in solution. This, in turn, is used in answering questions regarding stability and folding of proteins in membrane. Through spectroscopy and simulations, we probe the energy landscape of host-guest pentapeptides (WLLLX, where X is a variable residue).

### University of Missouri

Graduate Research Assistant

Oct 2019 - July 2024

- Visualization of SecDF: Applied AFM imaging and kymograph (line-scan) analysis to probe the conformational dynamics of protein translocation factor SecDF in the presence and absence of the Sec translocon SecYEG. Comparison to crystal structures and molecular dynamics simulations help bolster confidence in real-time, single-molecule data. These experiments suggest that at a basal level, the conformational dynamics of SecDF change in the presence of SecYEG.
- Chain-length Dependence on Peptide-Lipid Interactions: Applied single molecule force spectroscopy to probe the energetics and k kinetics of various peptides in the presence of a lipid membrane using approach/retract experiments. Peptides consisted of polyleucine constructs ( $WL_n$ , where n = 1...5), which are sufficiently small to not adopt higher order structures. This allows us to probe the effect of primary structure changes, namely length. Rupture force distributions were collected and analyzed, allowing us to determine the force-dependent dissociation rate, as well as the free energy profile. These parameters are typically hidden from traditional bulk techniques.

# EQUIPMENT AND SKILLS SUMMARY

- Programming Languages: IgorPro, TCL (For VMD/NAMD), Python, MATLAB, Java
- Cypher S Atomic Force Microscope: Tapping Mode Atomic Force Microscopy in Liquid, Force Spectroscopy
- Molecular Dynamics Software: VMD, NAMD, GROMACS
- FEI Scios DualBeam: Coupled Scanning Electron Microscope/Focused Ion Beam, AFM Cantilever Modification
- FEI Quanta 600F: Environmental Scanning Electron Microscope, X-Ray Energy Dispersive Spectroscopy

## RESEARCH AWARDS

- University of Missouri:
  - o Physics Leaders Award: Outstanding Presentation of Graduate Research 2nd Place Spring 2022
  - o Inaugural Chandrasekhar Fellow Spring 2024

## ACADEMIC AWARDS

- University of Missouri:
  - $\circ~$  Ernst Landen Fellowship Summer 2021
  - $\circ\,$  Anderson Physics Graduate Award Summer 2021
  - o Sigma Pi Sigma Student Inductee 2018

### Professional Organizations

- University of Wisconsin Eau Claire:
  - o Early Career Eau Claire 2024 to Present
- University of Missouri:
  - o Physics & Astronomy Grad Student Association: Chair of Educational Communication 2022 Calendar Year
  - o Physics & Astronomy Grad Student Association: Chair of Teaching Excellence 2021 Calendar Year
  - o PhysAssist Mental Health Counseling Graduate Student Mentor 2022 to 2024

### **Publications**

- [1]: R.S. Smith\*, **D.R. Weaver**\*, I. Kosztin, and G.M. King, Chain-length dependence of peptide-lipid bilayer interaction strength and binding kinetics: a combined theoretical and experimental approach, **Langmuir** 40, 28, 14467-14475 (2024). DOI: 10.1021/acs.langmuir.4c01218. (\*) equal contribution
- [2]: D.R. Weaver\*, K.G. Schaefer\*, and G.M. King, Atomic force microscope kymograph analysis: a case study of two membrane proteins, Methods 223, 83 (2024). DOI: 10.1016/j.ymeth.2024.01.013. (\*) equal contribution
- [3]: D.R. Weaver and G.M. King, Atomic force microscopy reveals complexity underlying general secretory system activity, International Journal of Molecular Sciences 24, 55 (2023). DOI: 10.3390/ijms24010055
- [4]: **D.R. Weaver**, D.N. Amin, and G.M. King, The conformations and basal conformational dynamics of translocation factor SecDF vary with translocon SecYEG interaction, **Journal of Biological Chemistry** 298, 102412 (2022). DOI: 10.1016/j.jbc.2022.102412

#### Presentations

- The Power of Touch: A Crash Course in Biological Atomic Force Microscopy: UWEC Physics Faculty Research Series. Authors: Dylan Weaver
- AFM-Based Force Spectroscopy Provides Quantitative Characterization of Single Residue Contributions in Peptide-Lipid Interactions: Graduate platform presentation at the 2024 Annual Meeting of the Biophysical Society (Feb 2024). Authors: Dylan Weaver, Ryan Smith, Ioan Kosztin, and Gavin King

#### Posters

- Probing the Effect of Secondary Structure in Peptide Binding and Detachment from Lipid Membranes using Melittin and its Diastereomeric Analog: Early career poster presentation at the 2025 Annual Meeting of the Biophysical Society (Feb 2025). Authors: Dylan Weaver, Ryan Smith, Krishna Sigdel, Milica Utjesanovic, Ioan Kosztin, and Gavin King
- Atomic Force Microscopy Kymographs Can Provide Robust Access to Conformational Transitions in Membrane Proteins: Graduate poster presentation at the 2023 Annual Meeting of the Biophysical Society (Feb 2023). Authors: Dylan Weaver, Katherine Schaefer, Divya Amin, Arthur Roberts, and Gavin King
- Direct Visualization of Conformations and Conformational Dynamics of the Proton-Driven Translocation Factor SecDF in Supported Lipid Bilayers: Graduate poster presentation at the 2022 Annual Meeting of the Biophysical Society (Feb 2022), and the 2022 Midwest Single Molecule Workshop (Aug 2022). Authors: Dylan Weaver, Divya Amin, and Gavin King
- Precise Analysis of Kymograph Data for Biological Atomic Force Microscopy using the Line Detection Algorithm: Graduate poster presentation at the 2021 Annual Meeting of the Biophysical Society (Feb 2021). Authors: **Dylan Weaver**, Lucas Chandler, and Gavin King

## University Service

- Early Career Eau Claire: A university group committed to providing resources and mentorship to early career (<6 years) faculty and staff at University of Wisconsin Eau Claire.
  - $\circ\,$  Core Planning Team: November 2024 Present
- Faculty Search Committees: Participated in a search to hire two new faculty for the Physics Department's new emphasis in Robotics Engineering.
  - $\circ$  2024 2025 Academic Year

### EXTERNAL SERVICE

- Journal Reviewer: Provided peer review for potential publications at the journal(s) listed below.
  - o Scientific Reports
- Biophysical Society Annual Meeting: Undergraduate Poster Award Competition (UPAC)
  - Volunteered with the Education Committee to judge undergraduate poster presentations and offer feedback for improvement.